



0040-4039(94)01636-4

## Relative Configuration of the C-1 to C-5 Fragment of Fumonisin B<sub>1</sub>

John W. ApSimon<sup>1</sup>, Barbara A. Blackwell<sup>2</sup>, Oliver E. Edwards<sup>1</sup> and Alain Fruchier<sup>3</sup>

<sup>1</sup>Ottawa-Carleton Chemistry Institute, Carleton University,  
Ottawa, Ontario, Canada, K1S 5B6.

<sup>2</sup>Plant Research Centre, Agriculture Canada,  
Ottawa, Ontario, Canada, K1A 0C6.

<sup>3</sup>École Nationale Supérieure de Chimie,  
34053 Montpellier Cedex 1, France.

**Abstract:** Synthesis of the 2,3-carbamate (**2**) and the 3,5-carbonate-N-p-bromobenzoate (**4**) derivatives of fumonisin B<sub>1</sub> have been made in an initial study of the configuration of fumonisins. These have been used to determine the relative configuration of the C(1)-C(5) fragment.

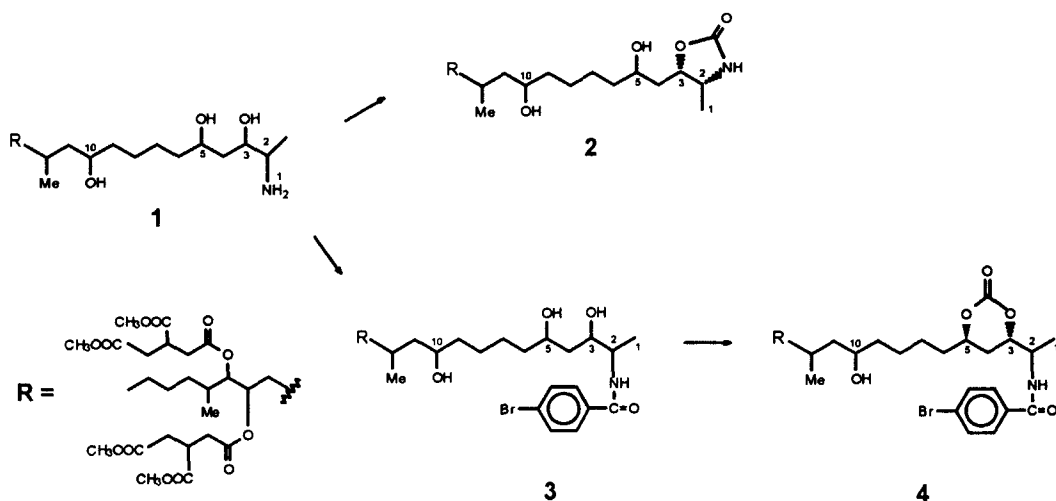
*Fusarium moniliforme* (Sheldon) is a prevalent mould on corn, sorghum and other grains throughout the world and has been shown to be toxic and carcinogenic for animals both as a contaminant of grains and as a pure culture.<sup>1,2</sup> This fungus contains a number of toxins, a group of which called fumonisins are thought to be mainly responsible for these diseases. Purified fumonisin B<sub>1</sub> (FB<sub>1</sub>) has been shown to cause equine leukoencephalomalacia,<sup>3</sup> porcine pulmonary edema<sup>4</sup> and to promote tumour formation in rats.<sup>5</sup> FB<sub>1</sub> has also been shown to be a potent inhibitor of ceramide synthetase,<sup>6</sup> thus blocking sphingolipid biosynthesis. This activity must be related to the stereochemical arrangement of the ten chiral centres in FB<sub>1</sub>. With the hope of obtaining a crystalline compound for X-ray analysis, a number of derivatives of FB<sub>1</sub> containing semi-rigid units were prepared. These failed to crystallize, but enabled assignment of the relative stereochemistry at carbons 2,3 and 5 using NMR spectroscopy.

**Results.** FB<sub>1</sub> tetramethyl ester (FB<sub>1</sub> Me<sub>4</sub>, **1**) was converted to the 2,3-carbamate (**2**) using triphosgene and triethylamine in benzene (25% yield).  $\nu_{\max}$  (CH<sub>2</sub>Cl<sub>2</sub>) 3600, 3530, 3445, and 1735 cm<sup>-1</sup>. FABMS,<sup>7</sup>  $m/z$  804 (M+1). <sup>1</sup>H and <sup>13</sup>C NMR,<sup>8</sup> see Tables 1 and 3.

The N-p-bromobenzoyl derivative **3** of compound **1** was prepared in 70% yield using p-bromobenzoyl chloride in a tetrahydrofuran/triethylamine mixture at 0°C.  $\nu_{\max}$  (CH<sub>2</sub>Cl<sub>2</sub>) 3600, 3520, 3425, 1735 (str), 1665 (med), and 1591 (wk) cm<sup>-1</sup>. FABMS,  $m/z$  960 (M+1). <sup>1</sup>H and <sup>13</sup>C NMR; see Tables 1 and 3.

The 3,5-carbonate **4** was prepared from **3** using phosgene in pyridine at 0°C (added Cr(OAc)<sub>3</sub> catalysed the reaction and prevented side reactions). Recycling twice was required to bring the yield to 60% based on unrecovered **3**.  $\nu_{\max}$  (CH<sub>2</sub>Cl<sub>2</sub>) 3520, 3420, 1738, 1655 cm<sup>-1</sup>. FABMS,  $m/z$  986 (M+1). <sup>1</sup>H and <sup>13</sup>C NMR; see Tables 1 and 3.

**Discussion.** Selected NMR data for compounds **1** to **4** are given in Tables 1 and 3, and are compared to the parent compound, FB<sub>1</sub>. Complete assignments have been made for all protons and carbons of **1-4**. These will be published elsewhere.<sup>9</sup> Chemical shifts varied little from those of FB<sub>1</sub>, except at the sites of substitution. The presence of the N-p-bromobenzoate ester induced a relatively large 1.4 ppm downfield shift of the H-2 resonance of **3**, whereas the formation of the carbamate ring induced a 0.8-0.9 ppm downfield shift. The introduction of the 3,5-carbonate ring in **4** caused only a 0.6 ppm shift in the relevant proton peaks.



Comparison of the  $^1\text{H}/^1\text{H}$  coupling constants of **2** and **4** is shown in Figure 1 and Table 2. Comparison of the carbonate coupling constants to those of the model compounds 2-oxo-1,3-dioxanes,<sup>10</sup> shows that the C-3 and the C-5 hydroxyl functions are trans in these cyclic compounds. The  $J_{3,4}$  value of 10.3 Hz in **4** is somewhat larger than that observed in  $\text{FB}_1$  (approximately 9 Hz) due to the fixed configuration induced by the ring. Analysis of the 2,3-carbamate spectrum shows that the C-2 amino function and the C-3 hydroxyl function are cis with respect to each other. The  $J_{2,3}$  of 6.5 Hz indicates a dihedral angle of approximately  $110^\circ$  or  $60^\circ$  between H-2 and H-3, depending on the relative orientation between H-2 and the terminal methyl. The  $110^\circ$  orientation is confirmed by the observation of an nOe effect between  $\text{CH}_3$ -1 and H-3. An amino-lipid with the same terminal stereochemistry has recently been reported.<sup>11</sup>

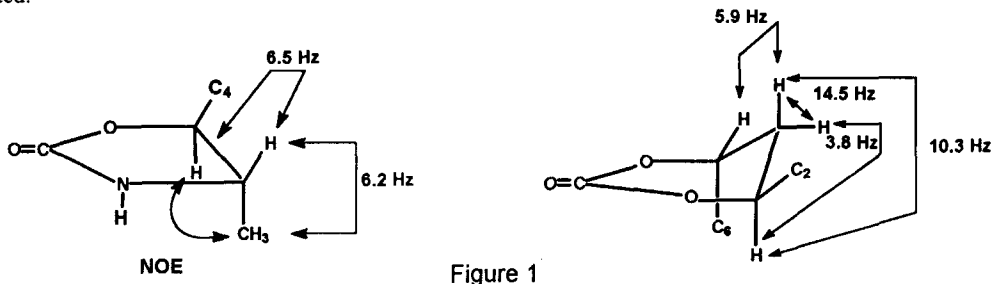


Figure 1

The  $^{13}\text{C}$  data are also consistent with the proposed stereochemistry. The chemical shifts are virtually constant except for C-1 to C-5. Conformational changes due to the bulky *p*-bromobenzoate group are likely accountable for some of the induced shifts. The formation of the carbonate (**4**) induces a 12 ppm upfield shift of C-4 and smaller downfield shifts of C-3 and C-5, consistent with the model compounds, the 2-oxo-1,3-dioxanes.<sup>10</sup> The presence of the carbamate and carbonate rings is confirmed by the presence of a carbonyl resonance at 158.6 ppm (**2**) and 149.3 ppm (**4**).

TABLE 1. Selected Proton Chemical Shifts in CDCl<sub>3</sub>\* of Fumonisin Derivatives 1-4 (ppm from TMS).

$\delta$	FB <sub>1</sub>	1	2	3	4
NH		-	5.21	6.63	6.36
H-1	1.27	1.06	1.26	1.30	1.43
H-3	3.14	2.80	3.57	4.17	4.49
H-3	3.74	3.48	4.38	3.98	4.55
H-4a	~1.55	1.43	1.63	1.57	1.95
H-4b		-1.63	1.74	1.73	2.13
H-5	3.84	3.84	3.88	3.92	4.48
H-10	3.62	3.57	3.60	3.61	3.55
H-14	5.16	5.15	5.17	5.19	5.18
H-15	4.94	4.88	4.90	4.91	4.90

TABLE 2. J<sub>H,H</sub> Coupling Constants of Fumonisin Derivatives 1-4 (Hz)

J <sub>H,H</sub>	FB <sub>1</sub>	1	2	3	4
NH, H-2	-	-	-	8.7	9.0
H-1, H-2	6.7	6.5	6.2	6.8	7.0
H-2, H-3	6.8	-	6.5	3.1	2.2
H-3, H-4a	3.2	-	3.1	3.1	3.8
H-3, H-4b	9.6	-	9.7	9.4	10.3
H-4 (AB)	-	-	-	14.6	14.6
H-4a, H-5	-	-	-	7.5	3.6
H-4b, H-5	-	-	-	3.5	5.9

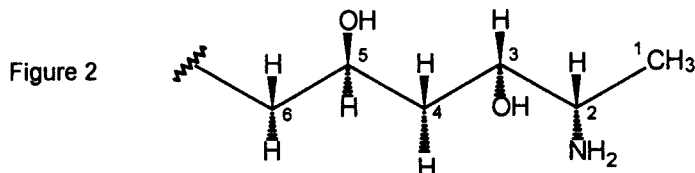
TABLE 3. Selected <sup>13</sup>C NMR Assignments of Fumonisin Derivatives 1-4 in CDCl<sub>3</sub>\*.

Position	FB <sub>1</sub>	1	2	3	4
1	16.06	20.10	20.15	18.30	17.89
2	53.76	51.23	53.93	50.19	47.52
3	70.43	73.21	81.29	71.63	78.09
4	41.83	39.75	41.58	40.25	28.49
5	68.48	68.49	67.73	69.26	77.38
6	39.01	37.56	38.00	37.18	34.56
10	69.98	68.82	68.80	68.37	68.72
14	72.80	71.26	71.10	71.22	71.06
15	78.82	77.82	77.83	77.86	77.83

\*FB<sub>1</sub> in CD<sub>3</sub>OD.

While biosynthetic studies show that the C-1, C-2 terminus is derived from l-alanine,<sup>12</sup> this does not define the absolute stereochemistry of this portion of FB<sub>1</sub>, since the asymmetric centre may not be preserved during decarboxylation. The relative stereochemistry of this terminus of the FB<sub>1</sub> molecule is as displayed in Figure 2. Comparison of the NMR data for the cyclic derivatives to those for FB<sub>1</sub> indicates that the relative configuration is the same as determined from the derivatives, which suggests that the fumonisin backbone forms a more rigid structure in solution than might be expected from this lipid-like molecule. This relative stereochemistry (Figure 2)

is opposite that of sphingosine (2,3 functionalities are threo). A recent study of AAL toxin<sup>13</sup> reports the same relative stereochemistry with respect to FB<sub>1</sub> between the adjacent amino and hydroxyl substituents, but the opposite stereochemistry for the hydroxyl function analogous to C-5 of FB<sub>1</sub>. In a parallel study, Poch et al.<sup>14</sup> have deduced the same relative conformation at C-2 and C-3.



*Acknowledgements.* We thank Pierre Lafontaine for the FABMS and John Nikiforuk for the NMR data. The cooperation of Drs. David Miller and Marc Savard in providing the FB<sub>1</sub> is also acknowledged. This work constitutes publication no. 1538 of Plant Research Centre.

#### REFERENCES AND NOTES

1. Gelderblom, W.C.A.; Jasliwka, K.; Marasus, W.F.O.; Thiel, P.G.; Horak, R.M.; Vleggaar, R.; Kriek, N.P.J.; *Carcinogenesis* **1988**, *9*, 1405.
2. Voss, K.A.; Plattner, R.D.; Bacon, C.W.; Norred, W.P.; *Mycopathologia*, **1990**, *112*, 81.
3. Thiel, P.G.; Marasus, W.F.O.; Sydenham, E.W.; Shephard, G.S.; Gelderblom, W.G.A.; *Mycopathologia*, **1992**, *117*, 3.
4. Harrison, L.R.; Colvin, B.; Greene, J.T.; Newman, L.E.; Cole, J.R.; *J. Vet. Diagn. Invest.*; **1990**, *2*, 217.
5. Gelderblom, W.C.A.; Kriek, N.P.J.; Marasus, W.F.O.; Thiel, P.G.; *Carcinogenesis*, **1991**, *12*, 1247.
6. Norred, W.P.; Wang, E.; Yoo, H.; Riley, R.T.; Merrill, A.H.; *Mycopathologia*, **1992**, *117*, 73.
7. **Fast Atom Bombardment Mass Spectrometry (FABMS).** Spectra were obtained on a Finnigan-MAT 312 spectrometer with an INCOS data system. The samples were introduced into the source as dispersions in thioglycerol.
8. **Nuclear Magnetic Resonance (NMR).** <sup>1</sup>H and <sup>13</sup>C NMR spectra were acquired on a Bruker AM500 spectrometer. Chemical shifts were referenced to deuteriochloroform at 7.24 ppm and 77.0 ppm for <sup>1</sup>H and <sup>13</sup>C respectively and are reported relative to tetramethyl silane (Me<sub>4</sub>Si). Chemical shift assignments were made with <sup>1</sup>H/<sup>1</sup>H (COSY) and <sup>1</sup>H/<sup>13</sup>C (HETCOR, HMBC) correlation spectra as well as DEPT and nOe experiments.
9. Blackwell, B.A.; Edwards, O.E.; Fruchier, A.; ApSimon, J.W.; in preparation for *Can. J. Chem.*
10. Pihlaja, K.; Rossi, K.; *Acta. Chem. Scand.*, **1993**, *B37*, 289.
11. Fanhua, K.; Faulkner, D.J.; *J. Org. Chem.*, **1993**, *58*, 970.
12. Blackwell, B.A.; Miller, J.D.; Savard, M.E.; *J. AOAC International*, **1994**, *77*, 506.
13. Oikawa, H.; Matsuda, I.; Ichihara, A.; Kohmoto, K.; *Tetrahedron. Lett.*, **1994**, *35*, 1223.
14. Poch, G.K.; Powell, R.G.; Plattner, R.D.; Weisleder, D.; *Tetrahedron Lett.*, accompanying communication.

(Received in USA 30 June 1994; revised 16 August 1994; accepted 23 August 1994)